

# Genome Sequences of *Gordonia terrae* Phages Attis and SoilAssassin

Welkin H. Pope, Daniel N. Biery, Zachary T. Huff, Amy B. Huynh, William M. McFadden, Julia S. Mouat, Scott E. Schneiderman, Hannah Song, Leah E. Szpak, Melanie S. Umbaugh, Brian A. German, Jill E. McDonnell, Nadia Mezghani, Claire E. Schafer, Paige K. Thompson, Megan C. Ulbrich, Victor J. Yu, Emily C. Furbee, Sarah R. Grubb, Marcie H. Warner, Matthew T. Montgomery, Rebecca A. Garlena, Daniel A. Russell, Deborah Jacobs-Sera, Graham F. Hatfull

Department of Biological Sciences, University of Pittsburgh, Pittsburgh, Pennsylvania, USA

**Attis and SoilAssassin are two closely related bacteriophages isolated on *Gordonia terrae* 3612 from separate soil samples in Pittsburgh, PA. The Attis and SoilAssassin genomes are 47,881 bp and 47,880 bp, respectively, and have 74 predicted protein-coding genes, including toxin-antitoxin systems, but no tRNAs.**

Received 9 May 2016 Accepted 18 May 2016 Published 30 June 2016

**Citation** Pope WH, Biery DN, Huff ZT, Huynh AB, McFadden WM, Mouat JS, Schneiderman SE, Song H, Szpak LE, Umbaugh MS, German BA, McDonnell JE, Mezghani N, Schafer CE, Thompson PK, Ulbrich MC, Yu VJ, Furbee EC, Grubb SR, Warner MH, Montgomery MT, Garlena RA, Russell DA, Jacobs-Sera D, Hatfull GF. 2016. Genome sequences of *Gordonia terrae* phages Attis and SoilAssassin. *Genome Announc* 4(3):e00591-16. doi:10.1128/genomeA.00591-16.

**Copyright** © 2016 Pope et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Welkin H. Pope, [welkin@pitt.edu](mailto:welkin@pitt.edu).

*Gordonia* spp. are implicated in foaming of sludge in wastewater treatment plants and are identified as opportunistic pathogens in hospital infections (1–4). Seventeen bacteriophages of *Gordonia* have been isolated, sequenced, and deposited in GenBank (5–9). It is unclear if the phages' genomic relationships reflect those of other phages of the phylum *Actinobacteria*, notably those of *Mycobacterium smegmatis* mc<sup>2</sup>155 whose phages exhibit a continuum of genetic diversity (10–16). The Science Education Alliance-Phage Hunters Advancing Genomics and Evolutionary Science (SEA-PHAGES) program is a course-based research experience in which undergraduates are immersed in research, using phage isolation and bioinformatics as a method to fuse authentic research and education (17). SEA-PHAGES has recently expanded its range of hosts for phage isolation to *Gordonia terrae* 3612.

Attis and SoilAssassin were isolated from separate soil samples at the University of Pittsburgh through direct plating of filtered soil extracts on *G. terrae* 3612. The phages were plaque purified and electron microscopy revealed that both phages have long non-contractile tails and isometric heads. DNA was isolated, and sequenced using Illumina MiSeq technology with 140 bp single-end reads. Reads were assembled using Newbler into major contigs for each phage of 47,881 bp and 47,880 bp with 949-fold and 932-fold coverages for Attis and SoilAssassin, respectively. Both have G+C% content of 66.8%, and discrete genome ends with 11-base single-stranded 3' extensions (5'-TACCAGGGGGA). BLASTn alignment of the two genomes shows that they differ by only a single base substitution and single bp insertion in Attis. Protein-coding genes were predicted using Glimmer (18), GeneMark (19), DNA Master (<http://cobamide2.bio.pitt.edu>), and Phamerator (20), and functions were assigned to 35 of the 74 genes in each genome using BLAST (21) and HHPred (22, 23) alignment against the publically available databases GenBank, the Protein DataBase, and pFamA. Predicted functions include those for virion structure genes, a tyrosine integrase and immunity repressor,

a RecET recombination system, RusA resolvase, and two HNH endonucleases.

Attis and SoilAssassin form plaques with distinct morphologies. SoilAssassin plaques are large (4-mm diameter), whereas Attis forms predominantly smaller plaques (1-mm diameter) although a few larger plaques are also observed. We predict that the single bp deletion at coordinate 25,254 of SoilAssassin in a putative minor tail protein gene is responsible for the plaque size difference. We note that Attis 29 likely corresponds to the parental form of the gene as the entire coding sequence is related to genes in several other distant phages. This relationship is reminiscent of the side tail fiber gene frameshift mutation in PaPa strains of phage lambda that facilitate large plaque formation relative to its Ur-lambda parent, which adsorbs to cells more rapidly than lambda PaPa thus reducing plaque size (24). Attis and SoilAssassin 29 genes are located at the 3' end of the virion structure and assembly operons, consistent with encoding tail fiber proteins.

Both phages encode putative tyrosine integrase and repressor genes near the center of their genomes, and are predicted to integrate into a tRNA<sup>ser</sup> gene corresponding to GTR9\_RS02590 in *Gordonia* sp. KTR9.

**Nucleotide sequence accession numbers.** The Attis and SoilAssassin genomes are available from GenBank under accession numbers [KU963247](https://www.ncbi.nlm.nih.gov/nuclot/KU963247) and [KU963246](https://www.ncbi.nlm.nih.gov/nuclot/KU963246), respectively.

## FUNDING INFORMATION

This work, including the efforts of Graham F. Hatfull, was funded by Howard Hughes Medical Institute (HHMI) (54308198).

## REFERENCES

- Blaschke AJ, Bender J, Byington CL, Korgenski K, Daly J, Petti CA, Pavia AT, Ampofo K. 2007. *Gordonia* species: emerging pathogens in pediatric patients that are identified by 16S ribosomal RNA gene sequencing. *Clin Infect Dis* 45:483–486. <http://dx.doi.org/10.1086/520018>.
- De los Reyes FL, III, Raskin L. 2002. Role of filamentous microorganisms in activated sludge foaming: relationship of mycolata levels to foaming initiation and stability. *Water Res* 36:445–459. [http://dx.doi.org/10.1016/S0043-1354\(01\)00227-5](http://dx.doi.org/10.1016/S0043-1354(01)00227-5).

3. De los Reyes FL, III, Rothauszky D, Raskin L. 2002. Microbial community structures in foaming and nonfoaming full-scale wastewater treatment plants. *Water Environ Res* 74:437–449. <http://dx.doi.org/10.2175/106143002X140233>.
4. Grisold AJ, Roll P, Hoenigl M, Feierl G, Vicenzi-Moser R, Marth E. 2007. Isolation of *Gordonia terrae* from a patient with catheter-related bacteraemia. *J Med Microbiol* 56:1687–1688. <http://dx.doi.org/10.1099/jmm.0.47388-0>.
5. Dyson ZA, Tucci J, Seviour RJ, Petrovski S. 2015. Lysis to kill: evaluation of the lytic abilities, and genomics of nine bacteriophages infective for *Gordonia* spp. and their potential use in activated sludge foam biocontrol. *PLoS One* 10:e0134512. <http://dx.doi.org/10.1371/journal.pone.0134512>.
6. Liu M, Gill JJ, Young R, Summer EJ. 2015. Bacteriophages of wastewater foaming-associated filamentous *Gordonia* reduce host levels in raw activated sludge. *Sci Rep* 5:13754. <http://dx.doi.org/10.1038/srep13754>.
7. Petrovski S, Seviour RJ, Tillett D. 2011. Prevention of *Gordonia* and *Nocardia* stabilized foam formation by using bacteriophage GTE7. *Appl Environ Microbiol* 77:7864–7867. <http://dx.doi.org/10.1128/AEM.05692-11>.
8. Petrovski S, Seviour RJ, Tillett D. 2011. Characterization of the genome of the polyvalent lytic bacteriophage GTE2, which has potential for biocontrol of *Gordonia*-, *Rhodococcus*-, and *Nocardia*-stabilized foams in activated sludge plants. *Appl Environ Microbiol* 77:3923–3929. <http://dx.doi.org/10.1128/AEM.00025-11>.
9. Petrovski S, Tillett D, Seviour RJ. 2012. Genome sequences and characterization of the related *Gordonia* phages GTE5 and GRU1 and their use as potential biocontrol agents. *Appl Environ Microbiol* 78:42–47. <http://dx.doi.org/10.1128/AEM.05584-11>.
10. Hatfull GF, Jacobs-Sera D, Lawrence JG, Pope WH, Russell DA, Ko CC, Weber RJ, Patel MC, Germane KL, Edgar RH, Hoyte NN, Bowman CA, Tantoco AT, Paladin EC, Myers MS, Smith AL, Grace MS, Pham TT, O'Brien MB, Vogelsberger AM, Hryckowian AJ, Wynalek JL, Donis-Keller H, Bogel MW, Peebles CL, Cresawn SG, Hendrix RW. 2010. Comparative genomic analysis of 60 mycobacteriophage genomes: genome clustering, gene acquisition, and gene size. *J Mol Biol* 397:119–143. <http://dx.doi.org/10.1016/j.jmb.2010.01.011>.
11. Jacobs-Sera D, Marinelli LJ, Bowman C, Broussard GW, Guerrero Bustamante C, Boyle MM, Petrova ZO, Dedrick RM, Pope WH, Science Education Alliance Phage Hunters Advancing Genomics and Evolutionary Science Sea-Phages Program, Modlin RL, Hendrix RW, Hatfull GF. 2012. On the nature of mycobacteriophage diversity and host preference. *Virology* 434:187–201.
12. Pope WH, Anders KR, Baird M, Bowman CA, Boyle MM, Broussard GW, Chow T, Clase KL, Cooper S, Cornely KA, DeJong RJ, Delesalle VA, Deng L, Dunbar D, Edgington NP, Ferreira CM, Weston Hafer K, Hartzog GA, Hatherill JR, Hughes LE, Ipapo K, Krukoni GP, Meier CG, Monti DL, Olm MR, Page ST, Peebles CL, Rinehart CA, Rubin MR, Russell DA, Sanders ER, Schoer M, Shaffer CD, Wherley J, Vazquez E, Yuan H, Zhang D, Cresawn SG, Jacobs-Sera D, Hendrix RW, Hatfull GF. 2014. Cluster M mycobacteriophages Bongo, PegLeg, and Rey with unusually large repertoires of tRNA isotypes. *J Virol* 88:2461–2480. <http://dx.doi.org/10.1128/JVI.03363-13>.
13. Pope WH, Bowman CA, Russell DA, Jacobs-Sera D, Asai DJ, Cresawn SG, Jacobs WR, Hendrix RW, Lawrence JG, Hatfull GF, Science Education Alliance Phage Hunters Advancing Genomics and Evolutionary Science, Phage Hunters Integrating Research and Education, >Mycobacterial Genetics Course. 2015. Whole genome comparison of a large collection of mycobacteriophages reveals a continuum of phage genetic diversity. *Elife* 4:e06416. <http://dx.doi.org/10.7554/eLife.06416>.
14. Pope WH, Ferreira CM, Jacobs-Sera D, Benjamin RC, Davis AJ, DeJong RJ, Elgin SC, Guilfoile FR, Forsyth MH, Harris AD, Harvey SE, Hughes LE, Hynes PM, Jackson AS, Jalal MD, MacMurray EA, Manley CM, McDonough MJ, Mosier JL, Osterbann LJ, Rabinowitz HS, Rhyan CN, Russell DA, Saha MS, Shaffer CD, Simon SE, Sims EF, Tovar IG, Weisser EG, Wertz JT, Weston-Hafer KA, Williamson KE, Zhang B, Cresawn SG, Jain P, Piuri M, Jacobs WR, Jr, Hendrix RW, Hatfull GF. 2011. Cluster K mycobacteriophages: insights into the evolutionary origins of mycobacteriophage TM4. *PLoS One* 6:e26750. <http://dx.doi.org/10.1371/journal.pone.0026750>.
15. Pope WH, Jacobs-Sera D, Best AA, Broussard GW, Connerly PL, Dedrick RM, Kremer TA, Offner S, Ogiefo AH, Pizzorno MC, Rockenbach K, Russell DA, Stowe EL, Stuke J, Thibault SA, Conway JF, Hendrix RW, Hatfull GF. 2013. Cluster J mycobacteriophages: intron splicing in capsid and tail genes. *PLoS One* 8:e69273. <http://dx.doi.org/10.1371/journal.pone.0069273>.
16. Pope WH, Jacobs-Sera D, Russell DA, Peebles CL, Al-Atrache Z, Alcoser TA, Alexander LM, Alfano MB, Alford ST, Amy NE, Anderson MD, Anderson AG, Ang AA, Ares M, Jr, Barber AJ, Barker LP, Barrett JM, Barshop WD, Bauerle CM, Bayles IM, Belfield KL, Best AA, Borjon A, Jr, Bowman CA, Boyer CA, Bradley KW, Bradley VA, Broadway LN, Budwal K, Busby KN, Campbell IW, Campbell AM, Carey A, Caruso SM, Chew RD, Cockburn CL, Cohen LB, Corajod JM, Cresawn SG, Davis KR, Deng L, Denver DR, Dixon BR, Ekram S, Elgin SC, Engelsen AE, English BE, Erb ML, Estrada C, Filliger LZ. 2011. Expanding the diversity of mycobacteriophages: insights into genome architecture and evolution. *PLoS One* 6:e16329. <http://dx.doi.org/10.1371/journal.pone.0016329>.
17. Jordan TC, Burnett SH, Carson S, Caruso SM, Clase K, DeJong RJ, Dennehy JJ, Denver DR, Dunbar D, Elgin SC, Findley AM, Gissendanner CR, Golebiewska UP, Guild N, Hartzog GA, Grillo WH, Hollowell GP, Hughes LE, Johnson A, King RA, Lewis LO, Li W, Rosenzweig F, Rubin MR, Saha MS, Sandoz J, Shaffer CD, Taylor B, Temple L, Vazquez E, Ware VC, Barker LP, Bradley KW, Jacobs-Sera D, Pope WH, Russell DA, Cresawn SG, Lopatto D, Bailey CP, Hatfull GF. 2014. A broadly implementable research course in phage discovery and genomics for first-year undergraduate students. *MBio* 5:e01051-01013. <http://dx.doi.org/10.1128/mBio.01051-13>.
18. Delcher AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying bacterial genes and endosymbiont DNA with glimmer. *Bioinformatics* 23:673–679. <http://dx.doi.org/10.1093/bioinformatics/btm009>.
19. Besemer J, Borodovsky M. 2005. GeneMark: web software for gene finding in prokaryotes, eukaryotes and viruses. *Nucleic Acids Res* 33:W451–W454. <http://dx.doi.org/10.1093/nar/gki487>.
20. Cresawn SG, Bogel M, Day N, Jacobs-Sera D, Hendrix RW, Hatfull GF. 2011. Phamerator: a bioinformatic tool for comparative bacteriophage genomics. *BMC Bioinformatics* 12:395. <http://dx.doi.org/10.1186/1471-2105-12-395>.
21. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *J Mol Biol* 215:403–410. [http://dx.doi.org/10.1016/S0022-2836\(05\)80360-2](http://dx.doi.org/10.1016/S0022-2836(05)80360-2).
22. Remmert M, Biegert A, Hauser A, Söding J. 2012. HHblits: lightning-fast iterative protein sequence searching by HMM-HMM alignment. *Nat Methods* 9:173–175. <http://dx.doi.org/10.1038/nmeth.1818>.
23. Söding J, Biegert A, Lupas AN. 2005. The HHpred interactive server for protein homology detection and structure prediction. *Nucleic Acids Res* 33:W244–W248. <http://dx.doi.org/10.1093/nar/gki408>.
24. Hendrix RW, Duda RL. 1992. Bacteriophage lambda PaPa: not the mother of all lambda phages. *Science* 258:1145–1148. <http://dx.doi.org/10.1126/science.1439823>.